

#### **RESEARCH ARTICLE**

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# **Behavioral alterations and neurotoxicity of Imidacloprid on freshwater Teleost** *Oreochromis mossambicus*

### Salunke Ankita<sup>1</sup>, Pandya Parth<sup>2</sup>, and Parikh Pragna<sup>1\*</sup>

<sup>1</sup>Department of Zoology, Faculty of Science, The Maharaja Sayajirao University of Baroda, Vadodara, India. <sup>2</sup>Division of Biomedical and Life Sciences, School of Sciences, Navarachana University, Vadodara, India. \*Corresponding author Email : <u>php59@vahoo.co.in</u>

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### Abstract

Agrochemicals are a significant cause of concern for the aquatic environment due to their toxicity, persistence and propensity to build up in the organisms. Imidacloprid (IMI) is a neonicotinoid insecticide widely used across the world. Improper and widespread usage and accidental exposure through agriculture runoff, IMI poses a major threat to nontarget aquatic organisms. Although fish being a major source of protein-rich food for the human being, concerns are raised against the health status and vulnerability of fish, leading to the entry of toxicants into the food chain. In this context, the present study is a replicating condition of insecticide exposure in the laboratory, where the toxic potential of IMI with their two sub lethal concentration (LC50/10th and LC <sub>50/20th</sub>) was tested (0.074 ppm and 0.04 ppm). Furthermore, the acute toxicity effect of IMI was validated by the behavioral response, histological alteration and biochemical estimation of Acetylcholine esterase (AchE) in the brain of Oreochromis mossambicus. Our result demonstrates that IMI has resulted in neuronal injury in the brain, resulting in severe abnormalities and suggests that alteration in AchE levels can be a good bioindicator for monitoring neuro-toxicity caused by insecticides.

**Keywords:** *Imidacloprid, sub-lethal, Oreochromis mossambicus, brain.* 

# Introduction

The potentially deleterious effect on various natural environment components has elevated a great deal of concern in scientific circles for pesticide management [1-2]. Due to low cost and broad-spectrum toxicity, it is estimated that more than 100,000 tons of pesticides have been applied in India, primarily agricultural pest control [3]. The increase in agricultural production, crop protection, and yield have increased due to the use of chemical pesticides. The abundant use of these chemicals, under the adage, "If little is good, a lot more will be better" has played an essential role in increasing the consumption [4-5]. The annual application of agricultural fertilizers and pesticides is over 140 billion kilograms, a vast source of pollutants through agricultural runoff [6-9]. Application of such agrochemicals shows potential health hazards and has become a major concern for aquatic habitat due to their toxicity, persistency and tendency to accumulate in the organisms. [10-12]. The application of pesticides has increased several folds in India and is likely to increase in the coming years. The applied insecticides into agriculture fields easily get washed away, enter into the aquatic system in vast quantities and imbalance the ecosystem and induce physiological and biochemical effects on aquatic organisms. Fishes are most essential and highest interacted species of the aquatic ecosystem. They become a bridge between aquatic and terrestrial ecosystems as they are getting used by birds and mammals as a food source. [13-16].

Over the last period, a new class of insecticides, the neonicotinoids, has developed the most important and fastest emerging pesticides on the global market [17-19]. When used as plant protection products, neonicotinoids becoming distributed function by systemically throughout the growing plant following seed or soil application. IMI is potential groundwater and surface water contaminant because it can leach and runoff from soil and crops [20-22]. It may also enter water bodies from spray drift or accidental spills, leading to local point-source contamination. There are reports which suggest that the minimum concentration of 10  $\mu$ g/L IMI in the aquatic environment may have adverse effects on the embryonic and larval stages of common carp. [23] and in L. variegatus by decreasing survival, inhibiting behavior, interfering with the growth process, and span [24]. Imidacloprid is a shortening life neonicotinoid insecticide that causes paralysis of the

central nervous system in insects. Moreover, it may also affect the central nervous system of humans and animals [25]. Previous work of our lab has also reported physiological, biochemical, and histopathological alteration in *O. mossambicus* in the exposure of IMI in liver, kidney, and gills. [9,26] However, there was a lacuna for the evaluation of histoarchitectural alteration and analysis of Acetylcholine esterase activity from the brain; hence the present work is an attempt to assess the neurotoxic potential of IMI regarding Acetylcholine Esterase activity and histological change in the brain of *O. mossambicus* on the exposure of IMI.

# Methodology

#### Animal maintenance

Mature Tilapia *Oreochromis mossambicus* ( $15\pm 2 \text{ cm}$ ,  $25\pm 1.9 \text{ g}$ ) of similar size in length and weight were acquired from the pure brooders of the Vadodara district and transferred to the laboratory. The acclimation period was 15 days at  $27 \pm 40^{\circ}$ C, pH 7.4  $\pm$  0.5, dissolved oxygen  $8 \pm 0.3 \text{ mg/L}$ , total hardness 188 mg/L CaCO3 with a 12:12 light: dark photoperiod. Fish were supplied daily with commercial fish food during acclimation. The maintenance of animal and experimental protocols were in accordance with the guideline of APHA-AWWA-WEF [27].

#### **Experimental protocol**

Based on the LC<sub>50</sub> value [9] of IMI sub-lethal study, where the toxic potential of IMI with their two sublethal concentrations LC  $_{50/10th}$  High Dose (HD) and LC  $_{50/20th}$  Low Dose (LD) was tested (0.074 ppm and 0.04 ppm). The experimental regime was maintained in the laboratory for 14 days, with a control group having three replicates in each group. The experiment was performed semi-statically with a group of 10 fishes (5 males and 5 females) in experimental aquaria. All the groups were kept under continuous observation during the experimental period for behavioural study like hyperactivity, restlessness, jerky movement, loss of equilibrium, fin movement and mucus secretion. After the completion of the exposure of 14 days, fishes were caught very gently using a small dip net, one at a time

with the least disturbance. They were slowly released in the tough containing 1% clove oil to make it immobile and afterwards allowed to dissect out the brain and was analysed by HE staining for its cellular architectural changes and histological alterations in fish exposed to low dose (LD) and high dose (HD) of IMI as compare to control. After measuring the weight, fresh tissues were fixed in 4% paraformaldehyde for 2 hrs, degraded and embedded in paraffin wax and sectioned at 10 -12 um then stained with haematoxylin and eosin and examined microscopically and photographed using a digital camera. After 14 days of exposure, the brains of Tilapia were homogenized in 0.1 M PBS. The homogenate was centrifuged at 10000 g at 4 °C for 15 min to obtain the supernatant. The supernatant was found to determine AChE activity. Triplicate samples from each treatment group were collected and measured the AChE activity by the Elman method 1961[28].

#### Statistical analysis-

The difference between the mean of control and the exposed fishes was determined by one-way ANOVA using Graph Pad Prism software version 6. If there was any significant difference, a post hoc test was carried out where Dunnett's multiple comparison tests were employed to recognize differences in the alterations that were found between the control and the exposed groups. The significant level of the tests was set at 5% (p < 0.05).

### **Results and Discussions**

#### **Behavioral Study**

Sub-lethal exposure of IMI resulted in an overall increase in behavioral patterns alteration compared to control. Frequency in the alteration of behavior, like restlessness, jerky movement, loss of equilibrium, and hyperactivity, was found to be more increased in HD than in LD treated IMI group and control. A significant change was observed in the tendency of fin movement and mucus secretion also. Based on this study, we can predict the alteration in behavior patterns is dose and exposure time dependent. (Table 1)

### **Histological Study**

A dose dependent alterations in the brain of O. mossambicus was observed on the exposure of IMI for a period of 14 days. Severity in histological changes was characterized by vacuolation (VC), necrosis of neurons (NC), with degeneration of neural cells, distended sinusoids (DS), and mild structural damages (SD) were observed in the brain of the fish exposed to a low dose of IMI. Whereas, when exposed to a high dose for a period of 14 days, significant changes were observed in the normal cytoarchitecture of the fish brain compared to control. Severe structural damage, increased necrosis, degenerative changes (DC), mononuclear infiltration (MI), congestion (C), cytoplasmic vacuolization (VC), and severe lesions (L) were also noticed in the brain. A significant haemorrhage (HE) was also noticed in some places. (Figure 1 and Table 2)

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Exposure time	Day	y1		Day 5			Day 1	0		Day 1	.5	
Dose	С	LD	HD	С	LD	HD	С	LD	HD	С	LD	HD
Jerky movement	+	+	+	+	+	+	+	+	++	+	+	++
Hyperactivity	-	-	-	-	+	+	-	+	++	-	+	+++
Restlessness	-	-	-	-	+	+	-	+	++	-	+	+++
Loss of Equilibrium	-	-	-	-	+	+	-	++	++	-	++	+++
Fin Movement	+	+	+	+	++	++	+	++	+++	+	++	+++
Mucus Secretion	-	-	-	-	+	+	-	+	++	-	++	++

Note: (-)  $\rightarrow$  Normal, (+)  $\rightarrow$  mild, (++)  $\rightarrow$  moderate, and (+++)  $\rightarrow$  maximum behavior pattern

Characterization	Control	LD	HD
Vacuolation	-	++	+++
Necrosis of neurons	-	++	+++
Distended sinusoids	-	++	+++
Degenerative changes	-	++	+++
Mononuclear infiltration	-	+++	+++
Congestion	-	++	+++
Lesions	-	+	+++
Hemorrhage	-	++	+++

Table 2: Summary of histological changes observed in the brain of *O. mossambicus* subjected to LD and HD of IMI

Note: (-)  $\rightarrow$  none, (+)  $\rightarrow$  mild occurance, (++)  $\rightarrow$  moderate occurrence, and (+++)  $\rightarrow$  maximum occurrence



LD

HD



Figure 1: Alteration in histology of brain in O. mossambicus on exposure of sub lethal dose of IMI



Figure 2: Acetylcholine Esterase Activity of the brain in O. mossambicus on exposure of sub lethal dose of IMI

#### Acetylcholine Esterase Activity

A Sublethal exposure of IMI resulted in a significant (p<0.05) decline of AChE activity, at high dose only as compared to control. (Figure 2)

## Discussion

In this study, the acute toxicity effect of IMI was studied in terms of alterations in the behavioral pattern, histopathological changes, and AChE activity in the brain of tilapia (*O. mossambicus*). The major aim was to validate the neurotoxic potential of IMI; the work was focused on alteration in behavior, AChE levels, and correlate it with the histoarchitectural changes in the brain.

Behavior provides a unique perspective on the physiology and ecology of an organism and its environment, operating through the central and peripheral nervous system [29]. Since behavior is not a random process, instead of a highly structured and anticipated sequence that ensures fitness and survival of the species, behavioral endpoints serve as a valuable tool to distinguish and evaluate the effects of exposure to environmental stress. Alteration in the fish behavior provides relevant indices for ecosystem assessment, any change in their behavior indicates the deterioration of water quality, so they are considered a biological indicator [30]. In the present scenario, the control fish were active and alert. They had a well-synchronized movement, whereas, in the case of IMI, exposure exhibited a dose-dependent severity in hyperactivity, loss of equilibrium, restlessness, and jerky movement. These symptoms may be due to inhibition of AChE activity leading to the accumulation of Ach in cholinergic synapses ensuing hyperstimulation. Since inhibition of AchE activity is a typical characteristic of neonicotinoids [31-32]; our findings corroborate with observations made by Rossi et al., 2020 in sentinel freshwater fish following sublethal exposure of neonicotinoids. The observed behavioral changes also suggest that the brain was affected by IMI exposure. To confirm these histological studies were conducted. Histology is used as a biomonitoring tool in toxicity studies as it provides early warning signs of disease. Histopathological observations have been commonly used to determine the health of the entire ecosystem population for toxicity testing of the effects of xenobiotic compounds at the sub-organismal or organismal level. [33-34]. The damage and injury in different organs are usually dependent on dose, duration, exposure, and type of pesticides. The utility of histological lesions is sensitive and reliable indicators of fish health as

reported in early studies. [20, 35]. Tissues obtained from the control group showed regular features with no apparent lesions. However, dose-dependent alterations in the brain were observed, such as Hyperplasia, edema, necrosis, vacuolation, and overall increase in brain cells. Vacuolation and increased cell size may have been due to glycolysis leading to microsomal and mitochondrial dysfunctions. [36-38]. The recent results of the present study show that the IMI causes major neurotoxicity in the brain and impairs its behavioral pattern. The problem can be more severe in the fish farms as the ponds are habitually located in or near the agricultural land, which is loaded with pesticides of all kinds. To this, if the groundwater used for domestic purposes can exacerbate the problem with simulated laboratory conditions. experimental Therefore, а scientific detoxification approach is necessary to improve the health of these economically valuable fish and reduce the losses caused by anthropogenic stress.

# Conclusion

The study suggests that sub-lethal concentration of IMI leads to neurotoxicity, and fishes were found to be highly stressed, therefore it is necessary to carry out field studies and check the fish health status and IMI residues in the environment. Furthermore, monitoring AChE levels can be used as a well-supported marker for IMI neurotoxicity in fishes in general and tilapia in particular.

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**Compliance with ethical standards** The research carried out is compliance with ethical standards.

### **Ethical approval**

All the procedures were in accordance with the guidelines of APHA-AWWA-WEF (1998) for which the care and use of animals were followed.

**Conflicts of interest:** The authors stated that no conflicts of interest.

# References

- Subash, S.P., Prem, C., Pavithra, S., Balaji, S.J. and Suresh, P. (2017). Pesticide Use in Indian Agriculture: Trends, Market Structure and Policy Issues.
- 2. Anand, M. and Taneja, A. (2020). Organochlorine pesticides residue in placenta and their influence on anthropometric measures of infants. *Environmental Research*. *182*, 106-109
- Sidhu, G.K., Singh, S., Kumar, V., Datta, S, Singh, D, Singh, J. (2019). Toxicity, monitoring and biodegradation of organophosphate pesticides: a review. Crit Rev Environ Sci Technol. <u>https://doi.org/10.1007/s00128990044</u>
- Patel, B., Pandya, P. and Parikh, P.H. (2016) Effect of agrochemicals on antioxidant enzymes and lipid peroxidation in *Oreochromis mossambicus* and *Labeo rohita*. *Journal of Zoology And Applied Biosciences*. 1(3):163-172.
- Pandya, P., And Parikh, P. (2016). Agrochemical induced gene expression alterations in *Oreochromis mossambicus*. *Int.J.Adv.Res.* 4(6):830-840.
- Dowd-Uribe, B., Press, D., Los Huertos, M. (2008). Agricultural Nonpoint Source Water Pollution Policy: The Case of California's Central Coast. Agriculture, Ecosystems & Environment. 128. 151-161. 10.1016/j.agee.2008.05.014.
- Ficklin, D., and Zhang, M.(2013). A Comparison of the Curve Number and Green-Ampt Models in an Agricultural Watershed. *Transactions of the ASABE*. 56. 61-69. 10.13031/2013.42590.
- Vymazal, J.,and Brezinova, T. (2014). The use of constructed wetlands for removal of pesticides from agricultural runoff and drainage: A review. *Environment International*. 75. 10.1016/j.envint.2014.10.026.
- 9. Patel, B., Upadhyay, A. and Parikh, P. (2016). Histological changes in the tissues of *Oreochromis* mossambicus and Labeo rohita on exposure to imidacloprid and curzate. International Journal of Research in Applied, Natural and Social Sciences, 4(5): 149-160.
- Pandya, P., Upadhyay, A., Thakkar, B., and Parikh, P. (2018). Evaluating the toxicological effects of agrochemicals on glucocorticoid receptor and serum cortisol level in Mozambique tilapia. *Cogent Biology*, 4(1), 1480338.

https://doi.org/10.1080/23312025.2018.1480338

- Karbalaei, S., Hanachi, P., Walker, T., Cole, M. (2018). Occurrence, sources, human health impacts and mitigation of microplastic pollution. *Environmental Science and Pollution Research*. 25. 36046–36063. 10.1007/s11356-018-3508-7
- Griffin, S., Love-Koh, J., Pennington, B., & Owen, L. (2019). Evaluation of Intervention Impact on Health Inequality for Resource Allocation. *Medical decision making: an international journal of the Society for Medical Decision Making*, 39(3), 171–182. <u>https://doi.org/10.1177/0272989X19829726</u>
- Sadekarpawar, S., Parikh, P. (2013). Gonadosomatic and hepatosomatic indices of freshwater fish *Oreochromis* mossambicus in response to a plant nutrient. World Journal of Zoology, 8: 110–118.
- Upadhyay, A., Pandya, P. and Parikh, P.H. (2014) Acute exposure of Pyrazosulfuron Ethyl induced Haematological and Blood Biochemical changes in the Freshwater Teleost fish Oreochromis mossambicus.Int.J.Adv.Res.Biol.Scie. 1(2):179-186.
- Sadekarpawar, S., Pandya P., Upadhyay, A. and Parikh P. (2015 a). A comparative assessment of trace metal accumulation in *Oreochromis mossambicus* and *labeo rohita* exposed to plant nutrient librel TM. *International Journal* of Current Advanced Research. 4(10): 441-449.
- Pandya, P., Ambegaonkar, A. and Parikh, P. (2020) Evaluating the toxic potential of agrochemicals on the hypothalamic-pituitary-thyroid axis in tilapia (*Oreochromis mossambicus*). Journal of Applied Ichthyology (36), 203-211 <u>https://doi.org/10.1111/jai.13998</u>
- Jeschke, P.and Nauen, R., Schindler, M. and Elbert, A. (2011). Overview of the Status and Global Strategy for Neonicotinoids. *Journal of agricultural and food chemistry*. 59. 2897-908.
- Casida, J. E., and Durkin, K. A. (2013). Neuroactive insecticides: targets, selectivity, resistance, and secondary effects. *Annual review of entomology*, 58, 99– 117. <u>https://doi.org/10.1146/annurev-ento-120811-153645</u>
- Kakoolaki, S., Sharifpour, I., Afsharnasab, M., Sepahdari, A., Mehrabi, M., Ghaednia, B., Nezamabadi, H. (2014). Effects of temperature on hematological and histopathological changes and survival rate of juvenile Fenneropenaeus vannamei experimentally challenged to White Spot Virus. *Iranian Journal of Fisheries Sciences*. 13. 91-102.
- Desai, B and Parikh, P. (2012). Impact of Curzate (fungicide) on Hematological Parameters of Oreochromis mossambicus. International Journal of Scientific & Engineering Research, 3(7): 1-6.
- 21. PAN Pesticides database, 2016

- 22. Tabassum, F., Mohan, J., and Smith, P. (2016). Association of volunteering with mental well-being: a lifecourse analysis of a national population-based longitudinal study in the UK. *BMJ open*, 6(8), e011327. <u>https://doi.org/10.1136/bmjopen-2016-011327</u>
- Islam, M.A., Hossen, M.S., Sumon, K.A. *et al.* Acute Toxicity of Imidacloprid on the Developmental Stages of Common Carp *Cyprinus carpio. Toxicol. Environ. Health Sci.* 11, 244–251 (2019). <u>https://doi.org/10.1007/s13530-019-0410-8</u>
- Sardo, A. M., and Soares, A. M. (2010). Assessment of the effects of the pesticide imidacloprid on the behaviour of the aquatic oligochaete Lumbriculus variegatus. *Archives of environmental contamination and toxicology*, 58(3), 648–656. <u>https://doi.org/10.1007/s00244-010-9470-0</u>
- Buszewski B., Bukowska, M., Ligor, M., Staneczko-Baranowska I. (2019). A holistic study of neonicotinoids neuroactive insecticides-properties, applications, occurrence, and analysis. *Environ Sci Pollut Res Int.* 26(34):34723-34740. doi:10.1007/s11356-019-06114
- Sadekarpawar, S., Desai, B. and Parikh, P. 2010. Acute Toxicity and Behavioural responses of Oreochromis mossmbicus (Peters, 1852) to Insecticide, Fungicide and Plant Nutrient. *Biohelica*. 1(2): 16-21.
- 27. APHA-AWWA-WEF. (1998) Book on standard methods for the examination of water and wastewater (20th ed.). National Government Publication.
- Ellman, G. L., Courtney, K. D., Andres, V. & Featherstone, R. M. ((1961) A new and rapid colorimetric determination of acetylcholinesterase activity. *Biochem. Pharmacol.* 7, 88–95
- Halappa, R. and David, M. (2009). Behavioural Responses of the Freshwater Fish, *Cyprinus carpio* (*Linnaeus*) Following Sublethal Exposure to Chlorpyrifos. *Turkish Journal of Fisheries and Aquatic Sciences*, 9, 233-238. http://doi.org/10.4194/trifas.2009.0218
- Renick, V. C., Weinersmith, K., Vidal-Dorsch, D. E., and Anderson, T. W. (2016). Effects of a pesticide and a parasite on neurological, endocrine, and behavioral responses of an estuarine fish. *Aquatic toxicology* (*Amsterdam, Netherlands*), 170, 335–343. https://doi.org/10.1016/j.aquatox.2015.09.010
- Lionetto, M.G., Caricato, R., Giordano, M.E., Pascariello, M.F., Marinosci, L. and Schettino, T. (2003). Integrated use of biomarkers (acetylcholinesterase and antioxidant enzymes activities) in *Mytilus galloprovincialis* and *Mullus barbatus* in an Italian coastal marine area. *Mar. Pollut. Bull.* 46(3): 324-330.
- 32. Tomizawa, M. and Casida, J. E. 2011. Neonicotinoid insecticides: highlights of a symposium on strategic molecular designs. *J Agric Food Chem.* 59: 2883–2886.

- Joseph, B., Raj, S. J., Edwin, B. T., Sankarganesh, P., Jeevitha, M. V., Ajisha, S. U., & Sheeja, S. R. (2010). Toxic effect of heavy metals on aquatic environment. *International Journal of Biological and Chemical Sciences*, 4, 939–952. doi:10.4314/ijbcs.v4i4.62976
- David, M. and Rao, K. (2015). Histopathological alterations in spleen of freshwater fish *Cyprinus carpio* exposed to sublethal concentration of sodium cyanide. *Open Veterinary Journal.* 5. 1-5.
- Sadekarpawar, S., Upadhyay, A., Mansoori, R. and Parikh, P. (2015b.) Biochemical and Oxidative Stress Response of Plant Nutrient LibrelTM In Oreochromis mossambicus and Labeo rohita. International Journal of Advanced Research. 3(5): 1587-1596
- Bose, M.T., Ilavazhahan, M., Tamilselvi, R. and Viswanathan, M. (2013). Effect of Heavy Metals on the Histopathology of Gills and Brain of Fresh Water Fish *Catla catla. Biomedical & Pharmacology Journal.* 6. 99-105. 10.13005/bpj/390.
- Rajini, A., Revathy,K. and Selvam,G. (2015) "Histopathological changes in tissues of *Danio rerio* exposed to sub lethal concentration of combination pesticide," *Indian Journal of Science and Technolog*, *8*,(18):1–12.
- Xavier, J., and Kripasana, K. (2020). Acute Toxicity of Leaf Extracts of *Enydra fluctuans* Lour in Zebrafish (*Danio rerio* Hamilton). *Scientifica*, 3965376.

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